

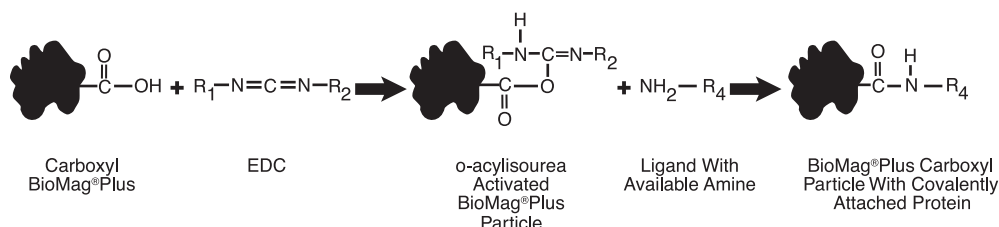
# BioMag®Plus Carboxyl & BioMag®Plus Carboxyl Protein Coupling Kit

## Description

BioMagPlus superparamagnetic microparticles are utilized in the magnetic separation of cells, organelles, proteins, immunoglobulins, nucleic acids, and many other types of molecules in biological and non-biological systems. The irregular shape of the BioMagPlus particle allows for much greater surface area, 20 to 30 times that of the same size spherical particle. This large surface area results in high binding capacities, allowing efficient target capture with minimal amounts of particles.

BioMagPlus consist of an iron oxide core with a silane coating. BioMagPlus particles are similar to conventional BioMag particles with the important distinctions of being much more uniform in size and more magnetically responsive. The more uniform size results in a more predictable and consistent behavior during capture and magnetic separation steps. Greater magnetic responsiveness results in faster magnetic separations especially on high throughput automated platforms.

BioMagPlus Carboxyl particles offer a high level of carboxyl functionality on magnetically responsive particles. Typically the carboxyl groups on the surface of the particles are activated by carbodiimide, allowing amine groups on proteins to be covalently attached.



Polysciences offers the BioMagPlus Carboxyl Protein Coupling Kit for covalently coupling proteins to BioMagPlus superparamagnetic particles. The contents of the kit are sufficient for five coupling reactions. To use the kit for smaller or larger samples adjust all volumes in a proportional manner.

## Characteristics

Mean Diameter: ~1 $\mu\text{m}$   
Particle Concentration: 20mg/ml

## Material Supplied

### Material Supplied with the BioMagPlus Carboxyl Protein Coupling Kit

Quantity	Kit Component	Volume
1	BioMagPlus Carboxyl	2.5ml
1	EDAC (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide)	0.10g
5	15ml conical centrifuge tubes	----
1	BioMag MultiSep Magnetic Separator	----
2	0.05M MES Buffer (pH 5.2)	175ml
1	Quenching Solution (1M Glycine, pH 8.0)	25ml
1	Wash Buffer	125ml

## Procedure

Researchers are advised to optimize the use of particles in any application. Activation and Protein Coupling steps should be performed in a well ventilated chemical fume hood.

### Activation

1. Transfer 0.5ml (10.0mg) of BioMagPlus Carboxyl particles to 15ml conical centrifuge tube. Magnetically separate the particles until the supernatant is clear. Aspirate the supernatant and discard.
2. Add 5ml of MES Buffer and shake well to mix. Magnetically separate the particles until the supernatant is clear. Aspirate the supernatant and discard.
3. Repeat Step 2, three times. After the final wash, resuspend the particles in 5ml of MES buffer.
4. Remove the EDAC from frozen storage and thaw at room temperature for a minimum of 30 minutes. Accurately measure out the required mass of EDAC (1.6mg EDAC/mg BioMagPlus Carboxyl), and add the EDAC to the particles in solution. Shake vigorously or vortex to mix.
5. Place the flask on a non-magnetic mixing device (rotator) for 30 minutes at room temperature. *Note:* The particles should not be allowed to settle during mixing.
6. Magnetically separate the activated particles until the supernatant is clear. Aspirate the supernatant and discard.
7. Repeat Step 2, four times.

### Protein Coupling

1. Calculate the amount of protein, antibody, or other ligand required for coupling. Generally, total protein concentrations of 20 to 500µg should be used for each milligram of activated BioMagPlus Carboxyl particles. Carrier proteins such as Bovine Serum Albumin (BSA) Fraction V (Cat. #23854) may be added to increase total protein concentration, blocking, and provide proper orientation of specific ligand binding for covalent attachment.
2. Add the protein to a volume of 5ml MES Buffer.
3. Remove 50µl of diluted protein solution and add it to 950µl of MES Buffer. This is a 1:20 dilution. Label as Pre-Coupling Solution. Set aside for Coupling Efficiency Determination.
4. Add the remaining protein solution (oligomer) to the activated particles and shake vigorously or vortex to mix. Place the centrifuge tube on a non-magnetic mixing device for 16 to 24 hours at room temperature.
5. Magnetically separate the particles until the supernatant is clear, and save. Label as Post-Coupling Solution. Set aside for Coupling Efficiency Determination.
6. Resuspend the particles in 5ml MES Buffer and shake vigorously or vortex to mix. Magnetically separate the particles until the supernatant is clear. Aspirate the supernatant and discard.
7. Repeat Step 5, one time.
8. Add 5ml of Quenching Solution. Shake vigorously or vortex to mix. Place the tube on a non-magnetic mixing device for 30 minutes at room temperature.
9. Magnetically separate the particles until the supernatant is clear. Aspirate the supernatant and discard.

### Washing and Diluting Coupled Particles

1. Add 5ml of Wash Buffer and shake vigorously or vortex to mix. Magnetically separate and discard supernatant.
2. Repeat Step 1, three times.
3. After the last wash, resuspend the particles to 2ml in Wash Buffer. Particle concentration is now approximately 5mg/ml.
4. Store the coupled BioMagPlus particles at 4°C as a suspension in Wash Buffer.

### Coupling Efficiency Determination

1. Set up a UV/Vis spectrophotometer to measure at 280nm. Fill both cuvettes with PWB and blank the spectrophotometer.
2. Measure the absorbance of the Pre-Coupling Solution and Post-Coupling Solution. *Note:* Further dilutions may be necessary to read the absorbance within the linear range of the instrument.
3. Calculate the coupling efficiency, expressed as the % Protein Uptake, as follows. Typical values of Protein Uptake are >60%.

$$\frac{[(A_{280} \text{ Pre-Coupling Solution} \times D) - (A_{280} \text{ Post-Coupling Solution} \times D)] \times 100}{(A_{280} \text{ Pre-Coupling Solution} \times D)}$$

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## Notes

1. Avoid use of amine (e.g. Tris) or carboxyl (e.g. acetate, citrate) buffers in the coupling step. Phosphate is satisfactory in the Coupling Buffer (i.e. prior to the attachment of protein). Buffers containing amine or carboxyl group can be used as Wash Buffers.
2. Some noncovalent adsorption invariably accompanies covalent coupling to particulate supports. Noncovalent adsorption is controlled by the washing procedure used after covalent protein attachment. The degree of noncovalent adsorption varies with each application and the washing procedure may have to be adjusted for individual applications. Additional washes to reduce noncovalently adsorbed protein can include high salt (1M NaCl), mildly acidic or basic media, mildly elevated temperatures, or increased time of exposure to the Wash Buffer. Dissociation of active, noncovalently adsorbed molecules from BioMagPlus particles can make magnetic materials appear unstable in some applications.
3. Prolonged vigorous shaking or vortexing can be used to resuspend BioMagPlus particles after magnetic separation or settling with gravity.

## Storage and Stability

Store at 4°C. Freezing, drying or centrifuging BioMag may result in irreversible aggregation and loss of binding activity.

**This product is for research use only and is not intended for use in humans or for *in vitro* diagnostic use.**

## Ordering Information

Cat. #	Description	Size
86011	BioMag®Plus Carboxyl	10ml
86010	BioMag®Plus Carboxyl Protein Coupling Kit	1 kit

### To Order

In The U.S. Call: 1-800-523-2575 • 215-343-6484

In The U.S. FAX: 1-800-343-3291 • 215-343-0214

In Germany Call: (49) 6221-765767

In Germany FAX: (49) 6221-764620

Order online anytime at [www.polysciences.com](http://www.polysciences.com).