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*Stain*RITE® May-Grünwald (MGG) Stain Procedure

1. Specimen - Peripheral smears should be prepared from a freshly drawn blood specimen *(EDTA purple top tubes)*. Allow smears to air dry completely before staining.

2. Please use an X or check mark to indicate that all stains and reagents are present on the bench before manual staining of rack. Reagents needed for manual testing are listed below.

REAGENTS CHECKLIST

_____ Methanol for ***Stain*RITE®** May-Grünwald Stain

_____ ***Stain*RITE®** May-Grünwald Stain

_____ ***Stain*RITE®** Giemsa Stain for MGG

_____ ***Stain*RITE®** Wright-Giemsa Buffer pH 6.8 or
***Stain*RITE®** MGG Buffer pH 7.2
(for darker blood smears)

_____ ***Stain*RITE®** Wright-Giemsa Buffer pH 6.8 or
***Stain*RITE®** MGG Buffer pH 7.2 with
***Stain*RITE®** May-Grünwald Stain Mixture

Please mix the ***Stain*RITE®** May-Grünwald Stain and buffer mixture before blood smears are made or before the smears are air drying.

Combine 45ml of ***Stain*RITE®** May-Grünwald Stain with 45ml of ***Stain*RITE®** Wright Stain Buffer pH 6.8 *(or the higher pH 7.2 buffer)*, mix well and allow to stand for at least 10 minutes. Make sure that a metallic sheen is observed on the surface before staining. Do not use after 2 hours of making the working buffer solution.

_____ ***Stain*RITE®** Giemsa Stain with Buffer Mixture

Combine 10ml of the G ***Stain*RITE®** iemsa Stain with 90ml of the ***Stain*RITE®** Wright-Giemsa Buffer Mixture or if a darker shade the ***Stain*RITE®** MGG 7.2 Buffer. Mix well and allow 10 minutes to stand before use. Use within 2 hours of mixing.

Allow to stand 10 minutes prior to use.

*Stain*RITE® MGG STAIN PROCEDURE

1. Place freshly prepared and air dried blood smears in a manual staining rack._____

2. Place slide rack with air dried blood smears in Methanol for 30 seconds. _____ *(Adjust timer for 30 seconds).*

3. Take a disposable pipette and flood the ***Stain*RITE®** May-Grünwald Stain on the appropriately labeled slides. _____

4. Set timer for 5 minutes. _____

5. Place oxidizing ***Stain*RITE®** May-Grünwald and ***Stain*RITE®** Wright-Giemsa Stain Buffer Mixture on ***Stain*RITE®** May-Grünwald stained slides laying on slide rack *(Displacing the **Stain*RITE®** May-Grünwald Stain off the slides with the pipette filled with **Stain*RITE®** May-Grünwald Stain/Buffer Mixture and viewing a metallic sheen on the top of slides).* _____

6. Set timer for 3 minutes. _____

7. Place slides in ***Stain*RITE®** Giemsa Stain/Buffer Mixture for 14 minutes.

8. Set timer for 14 minutes._____

9. Rinse slides in ***Stain*RITE®** Wright-Giemsa Stain Buffer or chosen buffer with appropriate pH for 2 minutes.

10. Retrieve slide rack with stained slides from buffer rinse and allow to air dry before examination with immersion oil and 100X objective.

STAINED SLIDE TIPS

1. Wipe excess stain from back of slides with methanol soaked gauze, **being careful not to wipe off the stained side of the slides.**
2. Place on slide envelope or slide flat/folder and proceed to the microscope room.
3. Place 1 drop of immersion oil onto slide to be viewed making sure it is

B. STAINING CHARACTERISTICS

Erythrocytes	light pink to moderate purple, not grey or blue
Polymorphonuclear Neutrophils	blue to dark blue to purple nuclei, reddish purple lilac granules, pale pink cytoplasm
Eosinophils	blue to dark blue to purple nuclei, red to orange-red granules, blue cytoplasm
Basophils	purple to dark blue to black nuclei, purple granules
Lymphocytes and monocytes	dark purple nuclei, sky blue cytoplasm
Platelets	violet to purple granules

ORDERING INFORMATION

Cat. #	Description
24981	***Stain*RITE®** May-Grünwald Stain Solution

Related Products

24984	***Stain*RITE®** Wright-Giemsa Stain Phosphate Buffer pH 6.8
24985	***Stain*RITE®** Wright-Giemsa Stain Solution
24986	***Stain*RITE®** Wright Stain Solution
24989	***Stain*RITE®** Wright Stain Phosphate Buffer pH 6.8
25032	***Stain*RITE®** May-Grünwald Giemsa Phosphate Buffer pH 7.2
25038	***Stain*RITE®** Giemsa Stain (for May-Grünwald)

TO ORDER

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