



**Instruction Manual** 

## **Purpose**

This study was conducted to test the validity of automated H & E staining, on routinely formalin fixed and paraffin embedded prostate chips, colon and kidney tissue.

## **Principle**

Differentiation is crucial for desired nuclear stain in regressive methods.

In progressive staining, immersion times are important to achieve good staining intensity.

## **Equipment**

- 1. Sakura Autostainer Prisma 81D 3. Leica Autostainer XL
- 2. Sakura Autostainer Coverslipper 4. Fisher Histomatic Autostainer

#### **Technique**

- 1. Paraffin sections were cut at 3 microns for slides with prostate chips, colon, and kidney samples
- 2. Slides with sections of colon, tonsil and placenta samples were cut at 4 microns
- 3. Skin samples were cut at 7 microns for frozen section
- 4. Slides were baked for 30 minutes in a 58°C oven

#### Reagents

- Xylene, Reagent Grade
- Alcohol, 95%, 100%
- 1N HCl to make 0.5% acid water
- Polysciences, Inc. Gill's #2 hematoxylin (cat. #24243)
- Polysciences, Inc. Gill's #3 hematoxylin (cat. #24244)
- Polysciences, Inc. Harris hematoxylin (cat. #24245)
- Polysciences, Inc. Eosin Y, 0.5% (cat. #09859)
- Polysciences, Inc. Eosin Y, 1.0% (cat. #17269)

- Polysciences, Inc. Ammonium Blue (cat. #24819)
- Polysciences, Inc. Lithium Blue (cat. #24820)
- Polysciences, Inc. Scott's Bluing Reagent (cat. #24605)
- Polysciences, Inc. Poly-Mount (cat. #08381)
- 0.045% Acid Alcohol (2600 ml of 70% alcohol and 1.2 ml of concentrated HCl)
- Ethyl alcohol
- 0.3% in-house ammonia water

# **Procedure for Automated Staining**

- 1. Deparaffinize slides to water
- 2. Stain sections on automatic stainer
- 3. Times for Polysciences' Harris and Gill's hematoxylin were both the same for regressive stains
- 4. Coverslip slides

# **Procedure for Manual Staining**

- 1. Fix frozen sections in 95% alcohol for 1 minute, then wash and air dry
- 2. Deparaffinize sections to water and air dry
- 3. Dip sections in water and stain in hematoxylin for 1 minute
- 4. Wash slides in water for about 20 dips
- 5. 10 dips in bluing reagent
- 6. Wash in water for 20 dips
- 7. Put slides in 95% alcohol for 10 dips
- 8. Treat slides with Eosin Y for 10 seconds
- 9. Dip 5-7 times in 95% alcohol
- 10. Two changes of absolute alcohol, 10 dips each
- 11. Two changes of xylene substitute, 10 dips each
- 12. Coverslip slides

#### Results

Stained nuclei appeared blue and cytoplasm was pink. Polysciences' Harris and Gill's hematoxylin needed 7-8 minutes and yielded good results. All Polysciences, Inc. bluing reagents demonstrated desirable results. Sections cut at 3 microns were stained lighter that those at 4-5 microns, which is ideal.

# **Table 1** Times for Sakura Autostainer - Prisma 81D

All buckets for the Sakura Autostainer - Prisma 81D hold 250 ml of reagent.

Step	Reagent	Polysciences' Harris/Gill's Regressive (time)	Polysciences' Gill's Progressive (time)
1	Wash	0:10	0:10
2	Hematoxylin	8:00	2:00
3	Wash	1:00	1:00
4	0.5% Acid Alcohol	0:01	_
5	Wash	1:45	_
6	Bluing	0:30	0:30
7	Wash	1:00	1:00
8	95% OH	0:30	0:30
9	Eosin	1:00	1:30
10	95% OH	0:10	0:10
11	100% OH	0:10	0:10
12	100% OH	0:30	0:30
13	100% OH	1:30	1:30
14	Xylene	1:00	1:00
15	Xylene	1:00	1:00
16	Exit Xylene	1:00 - 3:00	1:00 - 3:00

# **Table 2** Times for Leica Autostainer XL

All buckets for the Leica Autostainer XL hold 450 ml of reagent.

Step	Reagent	Polysciences' Harris Regressive (time)	Polysciences' Gill's #2, #3 Progressive (time)
1	Wash	0:10	0:10
2	Hematoxylin	8:00	2:00
3	Wash	1:00	1:00
4	0.045% Acid Alcohol	0:01	_
5	Wash	1:10	_
6	Bluing	0:30	0:30
7	Wash	1:00	1:00
8	95% OH	0:30	0:30
9	Eosin	2:30	2:30
10	95% OH	0:07	0:10
11	95% OH	0:10	0:10
12	100% OH	0:30	0:30
13	100% OH	1:00	1:00
14	100% OH	_	_
15	Xylene	1:00	1:00
16	Xylene	0:30	0:30
17	Exit Xylene	1:00 - 3:00	1:00 - 3:00

# **Table 3** Times for Fisher Histomatic Stainer

All buckets for Fisher Histomatic Autostainer hold 600 ml of reagent.

Step	Reagent	Polysciences' Harris Regressive (time)	Polysciences' Gill's #2, #3 Progressive (time)
1	Wash	0:10	0:10
2	Hematoxylin	8:00	2:00
3	Wash	0:50	0:50
4	0.045% Acid Alcohol	0:01	_
5	Wash	2:00	_
6	Bluing	0:20	0:20
7	Wash	1:00	1:00
8	95% OH	0:30	0:30
9	Eosin	2:00	3:00
10	95% OH	0:10	0:10
11	100% OH	0:30	0:30
12	100% OH	1:00	1:00
13	Xylene	1:00	1:00
14	Xylene	1:00	1:00

**Table 4**Nuclear and Cytoplasmic Staining Intensity on the Sakura Autostainer - Prisma 81D

Hematoxylin	Acid Alcohol	Blue	Eosin	Comments
Polysciences' Harris	0.045%	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Good
Polysciences' Harris	0.045%	Polysciences' Lithium Blue	Polysciences' Eosin Y	Good
Polysciences' Harris	0.045%	Polysciences' Scott's Bluing	Polysciences' Eosin Y	Excellent, provided good contrast
Polysciences' Gill's #3	_	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Excellent
Polysciences' Gill's #3	_	Polysciences' Scott's Bluing	Polysciences' Eosin Y	Average
Polysciences' Gill's #3	_	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Good, no bluish tinge
Polysciences' Gill's #3	_	Polysciences' Lithium Blue	Polysciences' Eosin Y	Good, no bluish tinge
Polysciences' Gill's #3	_	Polysciences' Scott's Bluing	Polysciences' Eosin Y	Good

**Table 5**Nuclear and Cytoplasmic Staining Intensity on Leica Autostainer XL

Hematoxylin	Acid Alcohol	Blue	Eosin	Comments
Polysciences' Harris	0.045%	Polysciences' Ammonium Blue	Polysciences' Eosin	Good combination with fresh acid alcohol
Polysciences' Harris	0.045%	Polysciences' Lithium Blue	Polysciences' Eosin	Good, crisp with fresh acid
Polysciences' Harris	0.045%	Polysciences' Scott's Bluing	Polysciences' Eosin	Good, crisp with fresh acid
Polysciences' Gill's	_	Polysciences' Ammonium Blue	Polysciences' Eosin	Good
Polysciences' Gill's	_	Polysciences' Lithium Blue	Polysciences' Eosin	Good, crisp with fresh acid
Polysciences' Gill's	_	Polysciences' Scott's Bluing	Polysciences' Eosin	Good, crisp stain

**Table 6**Nuclear and Cytoplasmic Staining Intensity on Fisher Histomatic Autostainer

Hematoxylin	Acid Alcohol	Blue	Eosin	Comments
Polysciences' Harris	0.045%	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Average
Polysciences' Gill's #2	_	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Good
Polysciences'	_	Polysciences'	Polysciences'	Excellent, crisp
Gill's #2		Lithium Blue	Eosin Y	stain
Polysciences'	_	Polysciences'	Polysciences'	Excellent, crisp
Gill's #2		Scott's Bluing	Eosin Y	stain
Polysciences'	_	Polysciences'	Polysciences'	Excellent, crisp
Gill's #2		Ammonium Blue	Eosin Y	stain
Polysciences'	_	Polysciences'	Polysciences'	Excellent, crisp
Gill's #2		Scott's Bluing	Eosin Y	stain

**Table 7**Nuclear and Cytoplasmic Staining Intensity with Manual Regressive / Progressive Stain

Hematoxylin	<b>Acid Alcohol</b>	Blue	Eosin	Comments
Polysciences' Harris	0.045%	Polysciences' Ammonium Blue	Polysciences' Eosin	Excellent
Polysciences' Harris	0.045%	Polysciences' Lithium Blue	Polysciences' Eosin Y	Excellent
Polysciences' Harris	0.045%	Polysciences' Scott's Bluing	Polysciences' Eosin Y	Excellent
Polysciences' Gill's #2, #3	_	Polysciences' Ammonium Blue	Polysciences' Eosin Y	Excellent
Polysciences' Gill's #2, #3	_	Polysciences' Lithium Blue	Polysciences' Eosin Y	Excellent
Polysciences' Gill's #2, #3	_	Polysciences' Scott's Bluing	Polysciences' Eosin Y	Excellent

This study was conducted by Sakina Sadiq, B.S., HT (ASCP), HTL (ASCP) for Polysciences, Inc.